

HYcheck™

HYcheck D/E Neutralizing Agar · HYcheck for Disinfection Control · HYcheck for Enterobacteriaceae · HYcheck Plate Count Agar With TTC · HYcheck for Total Count · HYcheck for Yeasts and Molds · HYcheck for Yeasts and Molds with TTC

Intended Use

HYcheck is a hygiene contact slide which is used for assessing the microbiological contamination of surfaces or fluids.

Summary and Explanation

Monitoring the microbial flora of environmental surfaces, walls, ceilings, and equipment is an important stage in achieving good manufacturing practices in factories handling foods, cosmetics or pharmaceuticals.^{1,2,3} To maintain good hygiene standards in hotels and restaurant kitchens, microbiological contamination must also be monitored.⁴ Methods to

monitor the environmental flora have been described using either swabbing techniques⁵ or contact plates.⁶ Contact slides were created to monitor the microbial flora of liquids (e.g. urine, milk) and equipment surfaces in the clinical and food industries.¹ Contact slides are statistically comparable to swab and contact plates for surface sampling.¹ The HYcheck contact slides were developed for the testing of fluids and surfaces for microbial cleanliness.

HYcheck is a double sided, hinged plastic paddle containing two agar surfaces. The agar surface extends above the paddle allowing for contact with test surfaces. The hinged paddle allows the agar surface to be easily held against each test area during sampling. The surface area of the paddle is clearly divided into seven units of one centimeter each to allow direct counting of microbial density per unit area.

The HYcheck range of hygiene control slides consists of seven media combinations designed to meet the various needs for monitoring different types of microbial contamination.

HYcheck D/E Neutralizing Agar has both sides coated with D/E Neutralizing Agar, a medium developed by Dey and Engley⁷ to neutralize a broad spectrum of disinfectants and preservative antimicrobial chemicals. D/E Neutralizing Agar neutralizes higher concentrations of residual antimicrobials, when compared with other standard neutralizing formulas, such as Lethen media, Thioglycollate media, and Neutralizing Buffer.^{8,9} Complete neutralization of disinfectants is important because disinfectant carryover can result in a false no growth result. D/E Neutralizing media effectively neutralize the inhibitory effects of disinfectant carryover,^{10,11} allowing differentiation between bacteriostasis and true bactericidal actions of disinfectant chemicals.

User Quality Control

Identity Specifications

HYcheck D/E Neutralizing Agar

Medium:	D/E Neutralizing Agar
Appearance:	Lavender
Microbial Limits Test:	Satisfactory
pH at 25°C:	7.6 ± 0.2

HYcheck for Disinfection Control

Media:	D/E Neutralizing Agar	Tryptic Soy Agar
Appearance:	Lavender	Light amber
Microbial Limits Test:	Satisfactory	Satisfactory
pH at 25°C:	7.6 ± 0.2	7.3 ± 0.2

continued on following page

HYcheck for Enterobacteriaceae

Media:	Violet Red Bile Glucose Agar	Tryptic Soy Agar
Appearance:	Reddish purple	Light amber
Microbial Limits Test:	Satisfactory	Satisfactory
pH at 25°C:	7.4 ± 0.2	7.3 ± 0.2

HYcheck Plate Count Agar with TTC

Medium:	Plate Count Agar with 0.01% TTC
Appearance:	Light amber
Microbial Limits Test:	Satisfactory
pH at 25°C:	7.0 ± 0.2

HYcheck for Total Count

Media:	Plate Count Agar	Plate Count Agar with 0.01% TTC
Appearance:	Light amber	Light amber
Microbial Limits Test:	Satisfactory	Satisfactory
pH at 25°C:	7.0 ± 0.2	7.0 ± 0.2

HYcheck for Yeasts and Molds

Media:	Rose Bengal Chloramphenicol Agar	Tryptic Soy Agar
Appearance:	Rose pink	Light amber
Microbial Limits Test:	Satisfactory	Satisfactory
pH at 25°C:	7.2 ± 0.2	7.3 ± 0.2

HYcheck for Yeast and Molds with TTC

Media:	Rose Bengal Chloramphenicol Agar	Tryptic Soy Agar with 0.01% TTC
Appearance:	Rose pink	Light amber
Microbial Limits Test:	Satisfactory	Satisfactory
pH at 25°C:	7.2 ± 0.2	7.3 ± 0.2

Cultural Response (approx inoculum 30-300 CFU)

HYcheck D/E Neutralizing Agar

Inoculate and incubate at 35 ± 2°C for 18-48 hours.

ORGANISM	ATCC*	GROWTH ON D/E AGAR
<i>Aspergillus niger</i> NCPF 2275	—	good
<i>Bacillus subtilis</i>	6633	good
<i>Candida albicans</i>	2091	good
<i>Escherichia coli</i>	25922*	good
<i>Pseudomonas aeruginosa</i>	27853*	good
<i>Staphylococcus aureus</i>	25923*	good
<i>Staphylococcus epidermidis</i>	12228*	good

HYcheck for Disinfection Control

Inoculate and incubate at 35 ± 2°C for 18-48 hours.

ORGANISM	ATCC*	GROWTH ON D/E	GROWTH ON TSA
<i>Aspergillus niger</i> NCPF 2275	—	good	good
<i>Bacillus subtilis</i>	6633	good	good
<i>Candida albicans</i>	2091	good	good
<i>Escherichia coli</i>	25922*	good	good
<i>Pseudomonas /nosa</i>	27853*	good	good
<i>Staphylococcus aureus</i>	25923*	good	good
<i>Staphylococcus epidermidis</i>	12228*	good	good

HYcheck for Enterobacteriaceae

Inoculate and incubate at 35 ± 2°C for 18-24 hours.

ORGANISM	ATCC*	GROWTH ON VRBGA	GROWTH ON TSA
<i>Enterobacter aerogenes</i>	13048*	good	good
<i>Enterococcus faecalis</i>	19433*	none to poor	good
<i>Escherichia coli</i>	25922*	good	good
<i>Proteus mirabilis</i> NCTC 11938	—	good	good
<i>Salmonella typhimurium</i>	14028*	good	good
<i>Shigella sonnei</i>	25931*	good	good
<i>Staphylococcus aureus</i>	25923*	none to poor	good

HYcheck Plate Count Agar with TTC

Inoculate and incubate at 35 ± 2°C for 18-24 hours.

ORGANISM	ATCC*	GROWTH ON PCA W/TTC
<i>Enterococcus faecalis</i>	19433*	good
<i>Escherichia coli</i>	25922*	good
<i>Proteus vulgaris</i>	13315	good
<i>Salmonella typhimurium</i>	14028*	good
<i>Staphylococcus aureus</i>	25923*	poor

HYcheck for Total Count

Inoculate and incubate at 35 ± 2°C for 18-24 hours.

ORGANISM	ATCC*	GROWTH ON PCA	GROWTH ON PCA W/TTC
<i>Enterococcus faecalis</i>	19433*	good	good
<i>Escherichia coli</i>	25922*	good	good
<i>Proteus vulgaris</i>	13315	good	good
<i>Salmonella typhimurium</i>	14028*	good	good
<i>Staphylococcus aureus</i>	25923*	good	poor

HYcheck for Yeasts and Molds

Inoculate and incubate at 30 ± 2°C for 18-48 hours.

ORGANISM	ATCC*	GROWTH ON RBCA	GROWTH ON TSA
<i>Aspergillus niger</i> NCPF 2275	—	good	good
<i>Candida albicans</i>	2091	good	good
<i>Escherichia coli</i>	25922*	none to poor	good
<i>Saccharomyces cerevisiae</i> NCYC 1211	—	good	good
<i>Serratia marcescens</i>	8100	none to poor	good
<i>Staphylococcus aureus</i>	25923*	none to poor	good
<i>Streptococcus pyogenes</i>	19615*	none to poor	good

HYcheck for Yeasts and Molds with TTC

Inoculate and incubate at 30 ± 2°C for 18-48 hours.

ORGANISM	ATCC*	GROWTH ON RBCA	GROWTH ON TSA W/TTC
<i>Aspergillus niger</i> NCPF 2275	—	good	good
<i>Candida albicans</i>	2091	good	good
<i>Escherichia coli</i>	25922*	none to poor	good
<i>Saccharomyces cerevisiae</i> NCYC 1211	—	good	poor
<i>Serratia marcescens</i>	8100	none to poor	good
<i>Staphylococcus aureus</i>	25923*	none to poor	poor
<i>Streptococcus pyogenes</i>	19615*	none to poor	good

The cultures listed are the minimum that should be used for performance testing.

*These cultures are available as Bactrol™ Disks and should be used as directed in Bactrol Disks Technical Information.

HYcheck for Disinfection Control has side one coated with D/E Neutralizing Agar (D/E) (see above), and side two coated with Tryptic Soy Agar (TSA). In 1955, Leavitt et al.¹² demonstrated that Tryptic Soy Agar supports excellent growth of both aerobic and anaerobic microorganisms. Tryptic Soy Agar is a general purpose medium that is recommended in multiple water and wastewater applications.¹³

HYcheck for Enterobacteriaceae has side one coated with Violet Red Bile Glucose Agar and side two coated with Tryptic Soy Agar, a general purpose growth medium. Violet Red Bile Glucose Agar is a selective medium used for the enumeration of *Enterobacteriaceae* in foods. Coliform bacteria have long been used as an index of fecal contamination in waters, and their presence in milk is used as an index of sanitation in milk processing.¹⁴ The presence of *Enterobacteriaceae*, coliforms, *Salmonellae*, *Klebsiella* or *Citrobacter*, in raw foodstuffs is an indicator of fecal contamination. Their presence after processing may indicate a failure in the manufacturing process.

HYcheck Plate Count Agar with TTC has both sides coated with Plate Count Agar with TTC (0.01% 2,3,5-Triphenyl Tetrazolium Chloride).

HYcheck for Total Count has side one coated with Plate Count Agar and side two coated with Plate Count Agar with 0.01% TTC. Plate Count Agar is used for enumerating bacteria in water, wastewater, food and dairy products.^{13,15-18} TTC is a redox indicator that is colorless in the oxidized form. TTC is reduced to insoluble triphenylformazan by certain actively metabolizing bacteria, resulting in a red color in the presence of bacterial growth.

There are two HYcheck products for yeasts and molds: 1) **HYcheck for Yeasts and Molds** has side one coated with Rose Bengal Chloramphenicol Agar and side two coated with Tryptic Soy Agar; 2) **HYcheck for Yeasts and Molds with TTC** has side one coated with Rose Bengal Chloramphenicol Agar and side two coated with Tryptic Soy Agar with 0.01% TTC. Rose Bengal Chloramphenicol Agar is recommended in the selective isolation and enumeration of yeasts and molds from environmental materials and foodstuffs. The pH of the medium is near neutrality for improved growth and recovery of acid sensitive strains.¹⁹⁻²¹

Principles of the Procedure

HYcheck D/E Neutralizing Agar

Tryptone provides carbon and nitrogen. Yeast Extract provides vitamins, cofactors and additional nitrogen and carbon. Dextrose provides fermentable carbohydrate. Sodium Thioglycollate neutralizes mercurials. Sodium Thiosulfate neutralizes iodine and chlorine. Sodium Bisulfite neutralizes formaldehyde and glutaraldehyde. Lecithin neutralizes quaternary ammonium compounds and Polysorbate 80 neutralizes phenols, hexachlorophene, formalin and, with lecithin, ethanol. Brom Cresol Purple is a colorimetric indicator. Bacto Agar is a solidifying agent.

HYcheck for Disinfection Control

D/E Neutralizing Agar (D/E) - side one

Tryptone provides carbon and nitrogen. Yeast Extract provides vitamins, cofactors and additional nitrogen and carbon. Dextrose provides

fermentable carbohydrate. Sodium Thioglycollate neutralizes mercurials. Sodium Thiosulfate neutralizes iodine and chlorine. Sodium Bisulfite neutralizes formaldehyde and glutaraldehyde. Lecithin neutralizes quaternary ammonium compounds and Polysorbate 80 neutralizes phenols, hexachlorophene, formalin and, with lecithin, ethanol. Brom Cresol Purple is a colorimetric indicator. Bacto Agar is a solidifying agent.

Tryptic Soy Agar (TSA) - side two

Tryptone and Soytone provide nitrogen, vitamins and minerals. The natural sugars from the soybean promote bacterial growth. Sodium Chloride maintains the osmotic balance of the medium. Bacto Agar is a solidifying agent.

HYcheck for Enterobacteriaceae

Violet Red Bile Glucose Agar (VRBGA) - side one

Yeast Extract provides vitamins, cofactors, nitrogen and carbon. Glucose provides a source of fermentable carbohydrate. Bacto Agar is a solidifying agent.

Tryptic Soy Agar - side two

Tryptone and Soytone provide nitrogen, vitamins and minerals. The natural sugars from the soybean promote bacterial growth. Sodium Chloride maintains the osmotic balance of the medium. Bacto Agar is a solidifying agent.

HYcheck Plate Count Agar (PCA) with TTC

Tryptone and Yeast Extract provide carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. TTC is a redox indicator. Bacto Agar is a solidifying agent.

HYcheck for Total Count

Plate Count Agar - side one

Tryptone and Yeast Extract provide carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. Bacto Agar is a solidifying agent.

Plate Count Agar with TTC - side two

Tryptone and Yeast Extract provide carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. TTC is a redox indicator. Bacto Agar is a solidifying agent.

HYcheck for Yeasts and Molds

Rose Bengal Chloramphenicol Agar (RBCA) - side one

Soytone provides carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. Rose Bengal and Chloramphenicol inhibit bacterial growth and restrict size and height of rapidly growing mold colonies. Bacto Agar is a solidifying agent.

Tryptic Soy Agar - side two

Tryptone and Yeast Extract provide carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. Bacto Agar is a solidifying agent.

HYcheck for Yeasts and Molds with TTC

Rose Bengal Chloramphenicol Agar - side one

Soytone provides carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. Rose Bengal suppresses bacterial growth and restricts size and height of rapidly growing mold colonies. Chloramphenicol inhibits bacteria. Bacto Agar is a solidifying agent.

Tryptic Soy Agar - side two

Tryptone and Yeast Extract provide carbon and nitrogen. Dextrose provides a source of fermentable carbohydrate. TTC is a redox indicator. Bacto Agar is a solidifying agent.

Precautions

1. Do not touch agar surface.
2. Do not use if there are signs of dehydration or contamination.

Storage

Store HYcheck slides at 2-15°C.

Expiration Date

The expiration date applies to the product in its intact container when stored as directed. Do not use a product if it fails to meet specifications for identity and performance.

Procedure

Materials Provided

(One type is provided per package.)

HYcheck D/E Neutralizing Agar

HYcheck for Disinfection Control

HYcheck for Enterobacteriaceae

HYcheck Plate Count Agar with TTC

HYcheck for Total Count

HYcheck for Yeasts and Molds

HYcheck for Yeasts and Molds with TTC.

Test Procedure

Surfaces

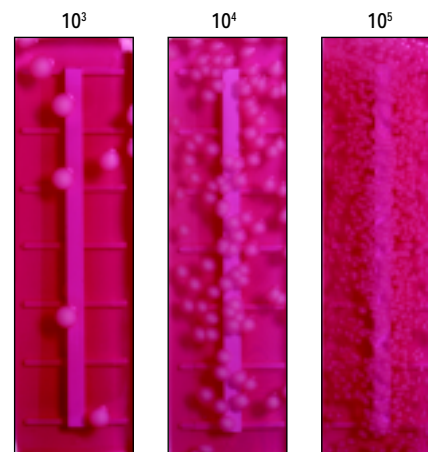
1. Loosen cap and remove HYcheck slide from the container.
2. Examine for dehydration or contamination.
3. Hold terminal spike against surface to be tested.
4. Press down on the spike to bend the paddle around the hinge line.
5. Gently lower the slide and press agar into contact with the test surface.
6. Apply firm and even pressure on the test surface for a few seconds.
7. Repeat procedure using the second agar surface on an area adjacent to the initial test site.
8. Replace slide in the container and close tightly.
9. Incubate in an upright position at indicated temperature.

Liquids

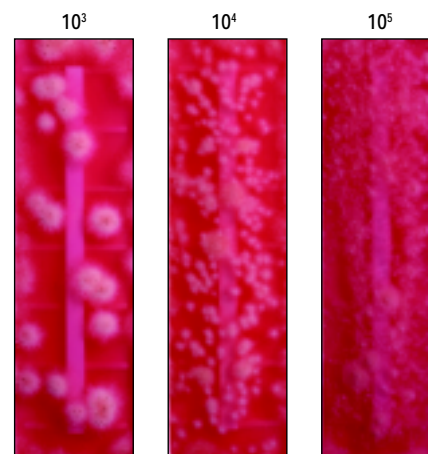
1. Loosen cap and remove HYcheck Slide from the container.
2. Examine for dehydration or contamination.
3. Immerse slide into test fluid so that agar surface becomes totally covered (if insufficient liquid is available, pour over surface of the slide).
4. Allow to drain.
5. Replace slide in the container and close tightly.
6. Incubate in an upright position at indicated temperature.

Results

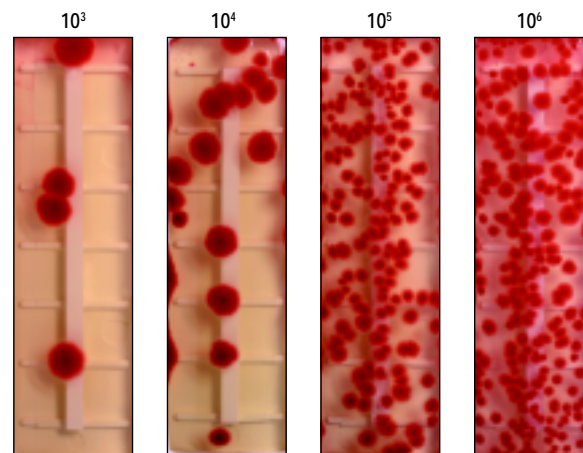
The following photos are reactions to *Candida albicans*, *Aspergillus niger* and *Escherichia coli*.



Candida albicans ATCC® 60193
on Rose Bengal Chloramphenicol Agar



Aspergillus niger ATCC® 1015
on Rose Bengal Chloramphenicol Agar



Escherichia coli ATCC® 11229
on Tryptic Soy Agar with 0.01% TTC

Limitations of the Procedure

1. Do not use the HYcheck Slide if it is contaminated or the agar medium is significantly dehydrated.

References

1. **Restaino, L.** 1994. HYcheck Slides versus contact plates compared to the swab technique. *Dairy, Food and Environ. Sanit.* **14**:528-530.
2. **Scott, E., S.F. Bloomfield, and C.G. Barlow.** 1984. A comparison of contact plate and calcium alginate swab techniques for quantitative assessment of bacteriological contamination of environmental surfaces. *J. Appl. Bact.* **56**:317- 320.
3. **Thomas, M. E. M., E. Piper, and I. M. Mauer.** 1972. Contamination of an operating theatre by Gram negative bacteria. Examination of water supplies, cleaning methods and wound infections. *J. Hygiene* **70**:63-73.
4. **Baird, R. M.** 1981. Cleaning and disinfection of the hospital pharmacy. S.A.B. Technical Series Number 16. Disinfectants: their use and evaluation of effectiveness.
5. **Griffiths, W. E.** 1978. Contact slides for use in environmental hygiene studies. *Environ. Health* **86**:36-37.
6. **Cain, R. M., and H. Steele.** 1953. The use of calcium alginate soluble wool for the examination of cleansed eating utensils. *Can. J. Pub. Health* **44**:464-467.
7. **Dey, B. P., and F. B. Engley, Jr.** 1970. A universal neutralizing medium for antimicrobial chemicals. Presented at the Chemical Specialties Manufacturing Association (CSMA) Proceedings 56th mid year.
8. **Dey, B. P., and F. B. Engley, Jr.** 1983. Methodology for recovery of chemically treated *Staphylococcus aureus* with neutralizing medium. *Appl. Environ. Microbiol.* **45**:1533-1537.
9. **Dey, B. P., and F. B. Engley, Jr.** 1978. Environmental sampling devices for neutralization of disinfectants. Presented at the 4th International Symposium on Contamination Control.
10. **Dey, B. P., and F. B. Engley, Jr.** 1994. Neutralization of antimicrobial chemicals by recovery media. *J. Microbiol. Methods* **19**:51-58.
11. **Dey, B. P., and F. B. Engley, Jr.** 1995. Comparison of Dey and Engley (D/E) neutralizing medium to letheen medium and standard methods medium for recovery of *Staphylococcus aureus* from sanitized surfaces. *J. Ind. Microbiol.* **14**:21-25.
12. **Leavitt, J. M., I. J. Naidorf, and P. Shugaevsky.** 1955. The undetected anaerobe in endodontics; a sensitive medium for detection of both aerobes and anaerobes. *The N.Y. J. Dentist.* **25**:377-382.
13. **Greenberg, A. E., L. S. Clesceri and A. D. Eaton (ed.).** 1995. Standard methods for the examination of water and wastewater, 19th ed. American Public Health Association, Washington, D.C.
14. **International Dairy Federation.** Milk and milk products-count of coliform bacteria. International Dairy Federation Standard FIL-IDF 73:1974.
15. **Swanson, K. J., F. F. Busta, E. H. Peterson, and M. G. Johnson.** 1992. Colony Count Methods, p.75-95. *In* C. Vanderzant, and D. F. Splittstoesser (ed.), Compendium of methods for the microbiological examination of foods, 3rd ed. American Public Health Association, Washington, D.C.
16. **Marshall, R. T. (ed.).** 1993. Standard methods for the examination of dairy products, 16th ed. American Public Health Association, Washington, D.C.
17. **Association of Official Agricultural Chemists.** 1995. Official methods of analysis, 16th ed. Association of Official Agricultural Chemists, Washington, D.C.
18. **Bandler, R., M. E. Stack, H. A. Koch, V. H. Tournas, and P. B. Mislivec.** 1995. Yeasts, molds and mycotoxins, p. 18.01-18.03. *In* FDA Bacteriological Manual, 8th ed. AOAC International, Arlington, VA.
19. **Martin, J. P.** 1950. Use of acid, rose bengal and streptomycin in the plate method for estimating soil fungi. *Soil Sci.* **69**:215-232.
20. **Koburger, J. A.** 1972. Fungi in foods. IV. Effect of plating medium pH on counts. *J. Milk Food Technol.* **35**:659-660.
21. **Jarvis, B.** 1973. Comparison of an improved rose bengal-chlortetracycline agar with other media for the selective isolation and enumeration of molds and yeasts in foods. *J. Appl. Bact.* **36**:723-727.

Packaging

HYcheck D/E Neutralizing Agar	20 units	9041-36
HYcheck for Disinfection Control	20 units	9039-36
HYcheck for Enterobacteriaceae	20 units	9037-36
HYcheck Plate Count Agar with TTC	20 units	9045-36
HYcheck for Total Count	20 units	9053-36
HYcheck for Yeasts and Molds	20 units	9038-36
HYcheck for Yeasts and Molds with TTC	20 units	9046-36